做科研，非一朝一夕

买器材，应速战速决

Newport数千种优质产品当日发货，更多惊喜尽在PhotonSpeed™光速购！
Measurement of algae PSII photosynthetic parameters using high-frequency excitation flashes

Chaoyi Shi (石朝毅), Yujun Zhang (张玉俊)*, Gaofang Yin (尹高方), Nanjing Zhao (赵南京), Jingbo Duan (段建波), Xiaoya Yu (余晓娅), Xue Xiao (肖 雪), Tingting Gan (甘婷婷), Li Fang (方 琳), and Wenqing Liu (刘文清)

State Environmental Protection Key Laboratory of Optical Monitoring Technology, Anhui Institute of Optics and Fine Mechanics, Chinese Academy of Sciences, Hefei 230031, China

*Corresponding author: yzhang@aiofm.ac.cn

Received February 25, 2014; accepted April 23, 2014; posted online July 18, 2014

We establish a system to measure the functional absorption cross section of photosystem II (PSII) (σ_{PSII}) and maximum quantum yield of photochemistry in PSII (F_m/F_o). The system utilizes a sequence of high-frequency excitation flashes at microsecond intervals to induce a microsecond-level fluorescence yield curve. Parameters σ_{PSII} and F_m/F_o are calculated by fitting the curve using nonlinear regression. Experimental results show that the relative standard deviation (RSD) of the system is less than 3%, and the correlation coefficient of F_m/F_o values measured by this system and those measured by pulse amplitude modulation method is 0.950.

OCIS codes: 010.4450, 120.0120, 300.6280.
doi: 10.3788/COL201412.080101.

Measurement of algae photosynthetic parameters provides theoretical basis for environmental pollution monitoring and algal bloom prediction, and helps to understand marine carbon cycle and marine primary productivity\cite{1-5}.

As a non-invasive, convenient, and reliable method, chlorophyll fluorescence method requires no extensive sample manipulation when measuring algae photosynthesis, thus it shows great potential for in situ measurement. This method has been well developed and widely used\cite{6-9}. Mauzerall\cite{10} first proposed the pump and the probe (P&P) fluorescence method for photosynthesis measurement. An intense pump flash is used to reduce all the primary electron acceptors Q_A, and the change of fluorescence yield is measured by a relatively low intensity probe flash. The probe flash intensity must low enough to avoid actinic effect, so the signal-to-noise ratio (SNR) is low in this method. Based on P&P method, Schreiber\cite{11} proposed pulse amplitude modulation (PAM) method, which permitted a high SNR by modulating the measuring light. But still, the modulation frequency must low enough (8–688 Hz) to avoid actinic effect, leading to a low time resolution. Consequently, this method cannot obtain a microsecond-level fluorescence yield curve, which allows the calculation of functional absorption cross section of photosystem II (PSII) (σ_{PSII}). The fast repetition rate (FRR) method put forward by Kolber and Falkowski provides a solution for the above problems\cite{12}. This method applies a sequence of excitation flashes at microsecond intervals to induce a microsecond-level fluorescence yield curve by reducing the primary electron acceptor Q_A, and parameters σ_{PSII} and F_m/F_o can be retrieved by fitting the induced curve. Based on the theory of FRR method, we established a PSII photosynthetic parameter measurement system using high-frequency excitation flashes at microsecond intervals, which permits the measurement of microsecond-level fluorescence yield curve and the calculation of σ_{PSII} and F_m/F_o.

In PSII, the quanta of excitation light are absorbed and transferred to reaction centers, and the overall efficiency of light trapping and energy transfer is described by the functional absorption cross section σ_{PSII}. The arrived excitation energy oxidizes reaction centre pigment P680 and reduces primary electron acceptor Q_A to Q_A, leading to the closure of PSII reaction center and then an increase of the fluorescence yield. Only after the reoxidation of Q_A (i.e., electron transfer from Q_A to plastoquinone (PQ) pool), can the reaction center reopen, and the fluorescence yield decline.

The fluorescence yield is determined by excitation energy, σ_{PSII}, and the redox state of Q_A. When excited by high-frequency excitation flashes at short enough intervals, all the Q_A can be reduced without any reoxidation. In such a situation, the fluorescence yield can be described as\cite{13,14}

$$F_i = F_o + (F_m - F_o) A_i = F_o + F_v \left[ 1 - \exp \left( -\sigma_{PSII} \sum_{j=1}^{i} l_j \right) \right],$$

where F_o is the minimal fluorescence yield measured when all PSII reaction centers are open, F_m is the maximum fluorescence yield measured when all PSII reaction centers are closed, F_v is variable fluorescence (i.e., the difference between the maximum and minimal fluorescence yield), A_i (0 \leq A_i \leq 1) is the fraction of PSII reaction centers closed at a given state of the excitation, and l_j is the energy of the jth flash.

To determine the photosynthetic parameters, we need to record the excitation flashes and the corresponding fluorescence signals, which are used to calculate the fluorescence yield curve. The fluorescence yield curve is then fitted into Eq. (1) using nonlinear regression to calculate σ_{PSII}, F_o, F_v, and F_m/F_o. σ_{PSII} is related to the type of
algae and the wavelength of the excitation light. In this letter, the discussion was limited to the measurements of \( \sigma_{\text{PSII}} \) for green algae (\textit{chlorella pyrenoidosa}) using 468-nm excitation LED.

The schematic of the measurement system is shown in Fig. 1. The system is composed of 468-nm LED array, LED driver unit, sample cell, detection unit for fluorescence signal and LED excitation light (used as reference signal), digital oscilloscope, and computer. To uniformly illuminate the sample cell, the 10 LEDs are arranged as a cycle around the sample cell, and low-pass glass filters are used to eliminate the long-wavelength radiation from the LEDs\(^{[15]}\). A microcontrol unit (MCU) is utilized to generate modulated signal with 2-\(\mu\)s duration at 1-\(\mu\)s intervals, which is then used to modulate the LEDs through a LED driver unit consist of TC4422A and MOSFET IRF640. The LEDs illuminate the sample cell with an average flux of 30000 \(\mu\)mol quanta/m\(^2\)/s, and the excitation energy is provided by 7 discharge capacitors.

The induced 685-nm fluorescence is collected from the bottom of the sample cell by a photomultiplier (PMT) with a 665-nm band-pass interference filter and a 600-nm long-pass glass filter\(^{[16]}\). The excitation light is detected by a PIN photodiode. The two signals are sampled by a digital oscilloscope with a sample rate of 20 MS/s and transferred to a computer. The fluorescence curve is calculated, and consequently the PSII parameters are retrieved on the computer.

The excitation light and stimulated fluorescence of healthy \textit{chlorella pyrenoidosa} sampled by the digital oscilloscope are shown in Figs. 2 and 3, respectively. The fluorescence yield is the ratio of the emitted fluorescence energy to the excitation energy absorbed by chlorophyll, but the measurement of the accurate absorbed excitation energy will greatly increase the complexity of the system and introduce more uncertain factors, hence the excitation energy is used instead. The integrations \( \text{LED}_i \) and \( \text{Fluo}_i \) are used as the energy of excitation flash and the energy of the corresponding fluorescence calculated from

\[
\text{LED}_i = \int_{t_i}^{t_i+\Delta t} l(t)dt, \tag{2}
\]

where \( \Delta t \) is the duration of the excitation flash. Consequently, the fluorescence yield is obtained from

\[
\text{Fluo}_i = \int_{t_i}^{t_i+\Delta t} f(t)dt, \tag{3}
\]

\[
F_i = \frac{\text{Fluo}_i}{\text{LED}_i}. \tag{4}
\]

Fig. 1. Schematic of the photosynthetic parameter measurement system using high-frequency excitation flashes.

Fig. 2. Excitation LED flash signal.

Fig. 3. Fluorescence signal corresponding to the excitation LED flashes.

Fig. 4. Fluorescence yield of healthy \textit{chlorella pyrenoidosa}.
The fluorescence yield $F_l$ is calculated as the ratio of the fluorescence energy $F_{0l}$ and the excitation energy $LED_l$, and $F_{0l}$ is in proportion to $LED_l$, thus $F_l$ is not influenced by both of the two intensity fluctuations, and in consequence $F_o$ and $F_v$ are both immune to the intensity fluctuations according to Eq. (1). But it is not the same for $\sigma_{PSII}$ because it is in inverse proportion to $\sum_j t_j$ as shown in Eq. (1). The intensity fluctuation between different measuring processes would do great harm to the accuracy of $\sigma_{PSII}$. The influence of this intensity fluctuation was eliminated by intensity correction. The excitation energy measured in the system calibration was used as a standard to correct this intensity fluctuation during the measurement. 8 parallel measurements of healthy *chlorella pyrenoidosa* were implemented and the results were shown in Table 1. The relative standard deviations (RSDs) of $\sigma_{PSII}$ and $F_v/F_m$ were respectively 2.96% and 1.62%, indicating a good stability.

Excessive Cu$^{2+}$ can damage the thylakoid membrane of algal cells, and inactivate the PSII reaction centers by inhibiting the electron transfer$^{[17,18]}$, which cause a change of the algae photosynthesis state. Therefore, thevalidity of the measurement system was analyzed by utilizing it to measure the photosynthetic parameters of the *chlorella pyrenoidosa* stressed by Cu$^{2+}$.

The concentrations of *chlorella pyrenoidosa* and Cu$^{2+}$ were respectively 100 $\mu$g/L and 25 $\mu$mol/L. The measurement was implemented every 5 min. With the increase of stress time, the rate of fluorescence yield saturation and the maximum fluorescence yield $F_m$ significantly declined (Fig. 5), and the retrieved $\sigma_{PSII}$ (Fig. 6) and $F_v/F_m$ (Fig. 7) also markedly declined.

The declines of $\sigma_{PSII}$ and the rate of fluorescence yield saturation indicated the damage of thylakoid membrane. As the light absorption and energy transfer were implemented on the thylakoid membrane, the damage of the thylakoid membrane caused the decline of $\sigma_{PSII}$, which resulted in a decrease of excitation energy arrived at reaction centers and in consequence a decline of the rate of fluorescence yield saturation.

The declines of $F_m$ and $F_v/F_m$ indicated the inactivation of PSII reaction centers. There was no photo-chemical action in inactivated reaction centers, and the arrived excitation energy dissipated as heat$^{[19]}$. Thus the fluorescence yield of the inactivated reaction centers remained $F_o$ even under ambient light, causing the decline of $F_m$ and consequently the decline of $F_v/F_m$.

As a comparison, the Water PAM instrument based on method was employed to measure $F_v/F_m^{[20]}$. The Water PAM can only measure $F_v/F_m$, but not $\sigma_{PSII}$. The correlation coefficient of the $F_v/F_m$ values measured by the system and those measured by Water PAM was 0.950 (Fig. 8). We also notice that the average value of $F_v/F_m$ measured by the system for healthy *chlorella pyrenoidosa* is 0.655, which is almost the same as the normal value (0.65)$^{[21]}$.

![Fig. 5. Fluorescence yield saturation curves with the stress time increasing from 0 to 60 min.](imageURL)

![Fig. 6. Measured $\sigma_{PSII}$ values of *chlorella pyrenoidosa* versus increasing stress time.](imageURL)

<table>
<thead>
<tr>
<th>No. of Samples</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\sigma_{PSII}$ ($\text{Å}^2$/quanta)</td>
<td>258.57</td>
<td>269.12</td>
<td>250.36</td>
<td>248.77</td>
<td>260.29</td>
<td>259.87</td>
<td>250.26</td>
<td>266.46</td>
</tr>
<tr>
<td>$F_v/F_m$</td>
<td>0.657</td>
<td>0.645</td>
<td>0.639</td>
<td>0.670</td>
<td>0.651</td>
<td>0.662</td>
<td>0.649</td>
<td>0.665</td>
</tr>
</tbody>
</table>
Fig. 7. Measured $F_v/F_m$ values of *chlorella pyrenoidosa* versus increasing stress time.

Fig. 8. (Color online) Correlation analysis of $F_v/F_m$ values measured by the established system and Water PAM in the Cu$^{2+}$ stress experiment.

In conclusion, we establish a PSII photosynthetic parameter measurement system using high-frequency excitation flashes. The system permits the induction of microsecond-level fluorescence yield curve, and calculates $\sigma_{PSII}$ and $F_v/F_m$ by fitting the fluorescence yield curve using nonlinear regression. The correlation coefficient of the $F_v/F_m$ values measured by this system and those measured by Water PAM method is 0.950. Furthermore, this system is extensible to induce fluorescence curves related to the electron transfer states of QA to PQ pool and PQ pool to PSI, revealing more photosynthesis details.

This work was supported by the Natural Science Foundation of Anhui Province (No. 1408085MD72), the National “863” Program of China (Nos. 2014AA06A509, 2013AA065502, and 2009AA063005), the Science and Technology Planning Project of Anhui Province (No. 1206c0805012), the National Natural Science Foundation of China (No. 61378041), and the Excellent Youth Foundation of Anhui Scientific Committee (No. 1108085J19).

References